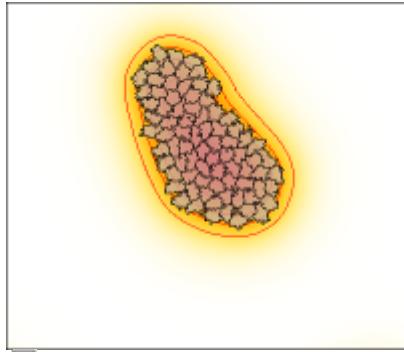
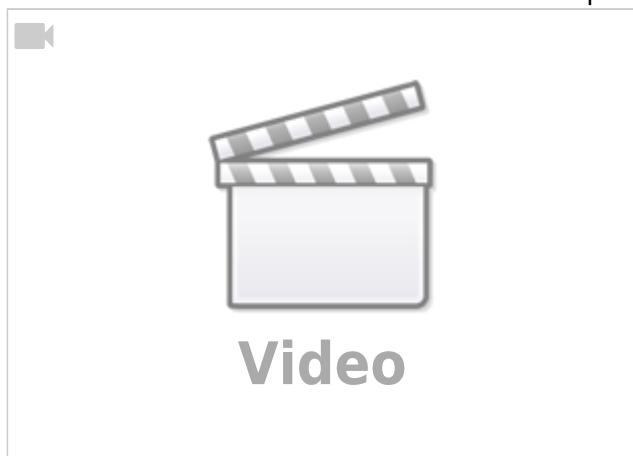


Multiscale models

Coupling spatial models: Autocrine chemotaxis



Cell cluster due to chemotaxis towards self-produced chemoattractant



Introduction

This example models a cellular Potts model for autocrine chemotaxis by coupling a CPM to a diffusive chemoattractant.

Model description

It exemplifies the combination of a CPM with a PDE through the chemotaxis towards a diffusive chemoattractant produced by cells. The `CellType` has a `Property p` (production rate) which is positive within biological cells, and is zero in the medium 'cell type'. The symbol `p` can then be used in the PDE as the production term for species or Layer `U`. In this way, `U` is produced in each lattice node that is occupied by a cell, and is not produced outside of cells. At the same time, cells perform Chemotaxis towards the chemoattractant `U`.

The simulation is visualized using some advances Gnuplotter feature. Cells are superimposed on the PDE and appear as transparent domains (see opacity). Moreover, the concentration gradient of the PDE is emphasized by the use of isolines that draw contours as positions with equal concentration.

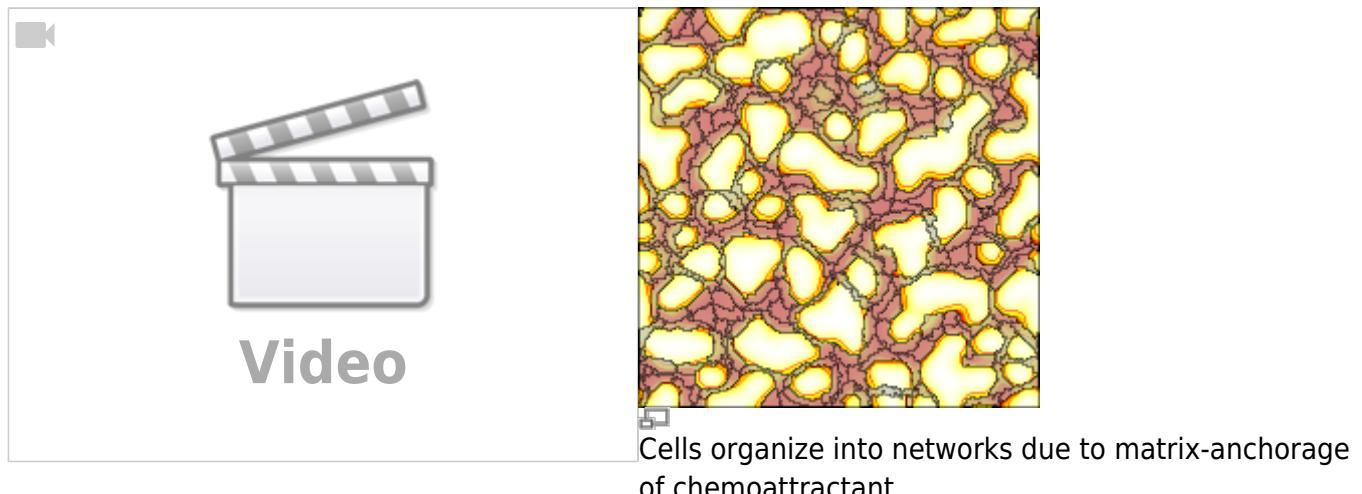
Full model

h AutocrineChemotaxis.xml |h

```
extern>
http://imc.zih.tu-dresden.de/morpheus/examples/Multiscale/AutocrineChemotaxis.xml
```

In Morpheus GUI: File → Examples → Multiscale → AutocrineChemotaxis.xml

Coupling CPM and PDE: Vascular patterning



Introduction

This example shows a model of vascular network formation by paracrine signaling (Köhn-Luque et al., 2011) and employs a coupled CPM and reaction-diffusion model.

Model description

The model defines a CPM as well as a PDE. These models are coupled by two processes:

1. Cells, specified in CellTypes, respond chemotactically to a Layer (or species) in the PDE
2. Conversely, the production term of one PDE Layer is coupled to the presence/absence of cell.

Full model

h VascularPatterning.xml |h

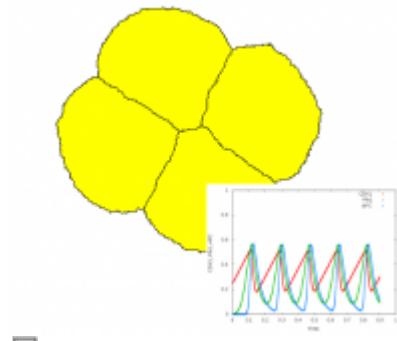
```
extern>http://imc.zih.tu-dresden.de/morpheus/examples/Multiscale/VascularPatterning.xml
```

In Morpheus GUI: File → Examples → Multiscale → VascularPatterning.xml

Reference

Köhne-Luque A, de Back W, Starruß J, Mattiotti A, Deutsch A, et al. [Early Embryonic Vascular Patterning by Matrix-Mediated Paracrine Signalling: A Mathematical Model Study](#). *PLoS ONE* 6(9): e24175, 2011

ODEs in CPM cells: Cell cycle and proliferation



Cells divide according to an oscillatory ODE model representing the early cell cycle in Xenopus.

Introduction

This multiscale model example shows

1. how to define a coupled system of continuous ODEs in discrete CPM cells
2. how to specify and change time scales between these model formalisms

Model description

This model specifies an oscillatory ODE model representing the cell cycle in Xenopus oocytes using three components (CDK1, Plk1, APK) (Ferrell et al., *Cell*, 2011) (see `CellTypes→CellType→System`). This ODE model is coupled to 2D shaped CPM cells that perform divide based on the concentration of these components (see `CellTypes→CellType→Proliferation→Condition`). As in the early Xenopus cell cleavage, this leads to exponential growth of the number of cells, without increase of total cell volume.

Time scales

Time scales are defined in the following fashion:

- The so-called global time scheme is defined in Time and here runs from 0 to 1 arbitrary time units. All models and plugins specify their updating scheme in terms this global time scheme (e.g. Analysis→Gnuplotter→interval).
- The CPM time scale for cell motility and behaviors is defined in CPM→MCSDuration. This specifies the time that a single Monte Carlo step in the CPM lasts, in terms of the 'global time'. Here, the MCSDuration is $1.0 \cdot 10^{-4}$ which means the CPM is executed 10.000 times during this simulation.
- For setting time of ODEs, one has to distinguish the (1) how often the ODEs are evaluated from (2) controlling the time scale of the ODE dynamics:
 1. One can set the time step between evaluation updates of the ODE System in System→time-step. This controls the *accuracy* of the numerical approximation (and is equal to the "ht" of the numerical solver).
 2. To change the time scale of the ODE dynamics, one needs to either change all the rate parameters or use a scaling prefactor (here v) to speed up or slow down make the dynamics.

Things to try

- Change the time scale of the CPM relative to the ODE dynamics: Increase CPM→MCSDuration to $1.0 \cdot 10^{-3}$ or decrease to $1.0 \cdot 10^{-5}$. This makes cells to have less resp. more motility/relaxation in between cell divisions.
- Change the time scale of the ODE dynamics: Increase CPM→MCSDuration to $1.0 \cdot 10^{-3}$ or decrease to $1.0 \cdot 10^{-5}$. This makes cells to have less resp. more motility/relaxation in between cell divisions.

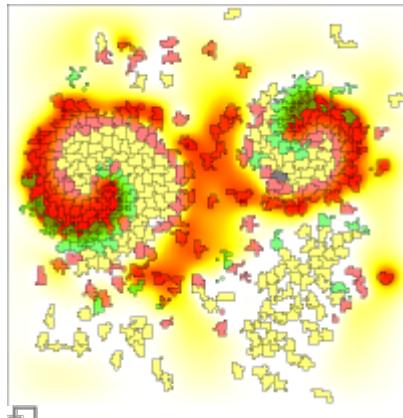
Full model

h CellCycle.xml |h

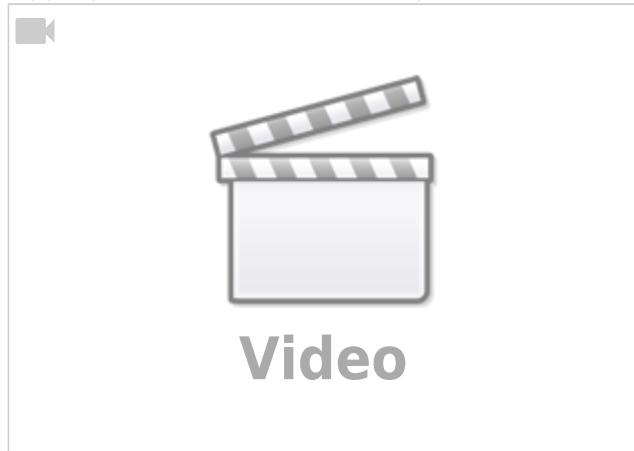
```
extern>
http://imc.zih.tu-dresden.de/morpheus/examples/Multiscale/CellCycle.xml
```

In Morpheus GUI: File → Examples → Multiscale → CellCycle.xml

Dictyostelium



Aggregation of amoebas through chemotaxis towards waves of cAMP.



Introduction

This model shows chemotactic aggregation of Dictyostelium. It was constructed by students attending the [ECMI modeling week 2012](#) in Dresden.

Model description

This model shows an interesting coupling between CPM cells and reaction-diffusion PDE. Cell state depends on the perceived concentration of cAMP, and determines whether a cell produces cAMP and whether it performs chemotaxis. The PDE is governed by a Fitzhugh-Nagumo-like model of an excitable medium, which causes traveling waves upon excitation. Chemotaxis through those waves causes cell aggregation.

Background colors indicate the cAMP concentration. Cells are color-coded according to their phase: excitable/resting (yellow), excited/chemotactic (green), refractory/resting (red).

Full model

h Dictyostelium.xml |h

```
extern>
```

<http://imc.zih.tu-dresden.de/morpheus/examples/Multiscale/Dictyostelium.xml>

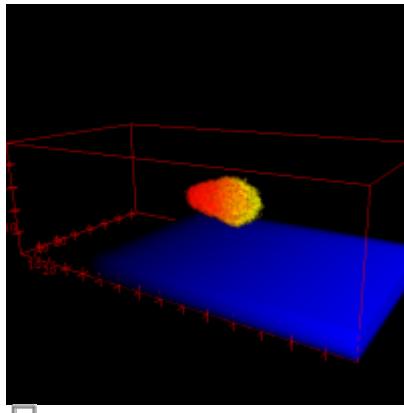
In Morpheus GUI: File → Examples → Multiscale → Dictyostelium.xml

References

- Savill N and Hogeweg P. [Modelling morphogenesis: from single cells to crawling slugs](#). *J. Theor. Biol.*, 184:229–235, 1997.
- Rost F, Quintero A, Myllykoski M, Igolkina A, Rohde O’Sullivan Freloft A, Dixit N, [Morphogenesis and Dynamics of Multicellular Systems](#). *ECMI Newsletter*, 52, 2012.

MembraneProperties: Cell polarization and chemotaxis

Note: MembraneProperties are not available in public version of Morpheus.



Cell dynamically re-polarizes in response to switching external gradient



Aims

This model shows how to couple cells, PDEs on membranes and external gradients.

Description

This example implements two models of cell polarity: Meinhardt's substrate-depletion model and Edelstein-Keshet's wave-pinning model. The user can switch polarity model by Disabling/Enabling the relevant System.

The model defines a one-dimensional reaction-diffusion system (MembraneProperty) representing membrane-bound molecules, and is mapped to a cellular Potts model defining a discrete shaped cell. An external gradient, specified in a PDE, provides a signal for the polarization of the cell. In turn, the polarity of the cell influences its chemotactic behavior.

After a switch in direction of the gradient, the cell re-polarizes in the new direction and starts to move up the gradient, iff the wave pinning model has been selected.

Full model

h CellPolarity.xml |h

```
extern>
http://imc.zih.tu-dresden.de/morpheus/examples/Multiscale/CellPolarity.xml
```

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